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# A facile procedure for the isolation of N-acetylneuramic acid from edible bird's-nest

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Sialic acids are commonly found as the nonreducing sugar residue in a variety of glycoproteins, mucins, and gangliosides which appear to play an important role in the plasma membrane of eucaryotic cells. N-Acetylneuraminic acid (1) is the best characterized sialic acid, and it can be prepared either by two tedious synthetic routes<sup>1,2</sup> or by isolation<sup>3-5</sup> from natural products available to laboratories with access to medical or veterinary sources. More recently, Schauer and Buscher<sup>b</sup> reported the isolation, for radiolabeling purposes of 1 from "edible bird's-nest". This procedure involved methanolysis, followed by ion-exchange chromatography which provided the methyl  $\beta$ -glycoside; 1 was then obtained by N-reacetylation followed by hydrolysis of the glycosidic linkage. When a hot-water extract of "edible bird's-nest" was studied as an inducer for the synthesis of neuraminidase from an Arthrobacter isolate<sup>7</sup>, it was observed that the extract from this commercial source contains large quantities of free sialic acids. We report, on this basis, a facile isolation procedure which reproducibly yields gram quantities of 1. The steps in the isolation procedure include a hot-water extraction, followed by two ultrafiltrations, and finally ion-exchange chromatography.

## **EXPERIMENTAL**

General methods. — Melting points were determined with a Kosler hot-stage, and correspond to corrected values. Optical rotations were measured with a Beckman 145 polarimeter. <sup>13</sup>C-N.m.r. spectra were recorded with a Varian XL-100-15 spectro-photometer operating at 25.16 MHz in the pulsed, Fourier-transform mode with complete proton-decoupling. Thin-layer chromatography (t.l.c.) was performed on cellulose plates (Eastman Kodak Co., Rochester, NY 14650, U. S. A.) with butanol-n-propanol-0.1M HCl (1:2:1, v/v) development; spots were located by spraying the plates with the Ehrlich reagent<sup>8</sup> followed by heating at ~110°. Alternatively, t.l.c. was performed with Silica Gel F 254 (Woelm, ICN Pharmaceuticals, 3340 Eschwege,

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West Germany) developed with *n*-propanol-water (7:3, v/v); spots were located by spraying the plates with 10%  $H_2SO_4$  and heating at ~110°.

Analysis of sugars. — The direct Ehrlich reaction was performed as described by Gottschalk<sup>9</sup>. Sialic acids were quantitatively determined with the thiobarbituric assay as described by Warren<sup>10</sup>.

Isolation of N-acetylneuraminic acid. — Pulverized "edible bird's-nest" (300 g) was added to boiling water (61), and the solution was boiled under reflux for 5 h. The solids were allowed to settle, and the supernatant ( $\sim 5$  l) was obtained by suction filtration. The hot-water bird's-nest extract was concentrated to  $\sim 750$  ml by partial freeze-drying. The solution (12.6 g of sialic acids) was subjected to a two-step ultra-filtration ( $N_2$ ), first through a Diaflo XM-50 filter (Amicon Corporation, Lexington MA 02173) followed by two washings with water and then through a Diaflo UM-10 filter followed by repeated water washings until free of sialic acids. Purification of 1 was achieved by ion-exchange chromatography of the combined filtrate and washings on a column of Dowex 1-X4, as shown in Fig. 1. The eluate from the column with formic acid was rapidly screened for sialic acids by the direct Ehrlich<sup>8</sup> assay, which

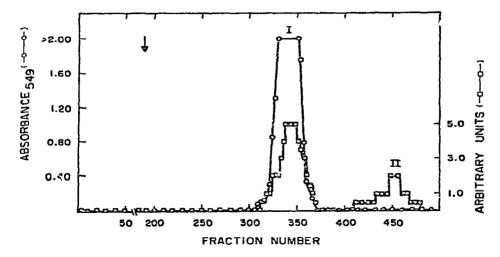


Fig. 1. Purification of 1 by ion-exchange chromatography. A column  $(4.5 \times 70 \text{ cm})$  of Dowey 1-X4 (HCO<sub>2</sub>) anion-exchange resin was prepared essentially as described by the distributor (BioRad, Richmond, CA 94804, U. S. A.). The DM-10 ultrafiltrate was added, followed by washing with water (~21). At the indicated point (arrow), a tinear formic acid gradient (41, 0-2M) was applied. The flow rate was 120 ml/h, and 20-ml fractions were collected. From every third fraction, a sample  $(5 \mu l)$  was examined by Ehrlich<sup>8</sup> (——) and Warren<sup>10</sup> (——) tests. For the former assay, the arbitrary units refer to relative color intensities.

revealed two peaks. Only Peak I consisted of material giving a positive Warren reaction <sup>10</sup>. The latter fractions were combined, and the formic acid was removed by Hash evaporation at a bath temperature <40°. The resulting residue was dissolved in water, and crude I was obtained by lyophilization. When necessary, final purification was accomplished by a second passage of the redissolved material through the

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same column under identical conditions. The pooled, lyophilized material was obtained in a final yield of 6.5 g. It proved to be homogeneous by t.l.c. on cellulose and Silica Gel F, and migrated at the same speed as a standard of 1 (synthetic, Sigma Chemical Co., St. Louis, MI 63178);  $[\alpha]_D^{20} - 31.7$  (c 2.0, water); lit.<sup>5</sup>:  $[\alpha]_D^{20} - 32^\circ$ . The lyophilized material was recrystallized from water-glacial acetic acid (1:10), m.p. 187-189° (dec.); lit.<sup>5</sup>: m.p. 187° (dec). The methyl ester of 1, homogeneous by t.l.c., was prepared as described by Kuhn et al.<sup>11</sup>, m.p. 182-184°,  $[\alpha]_D^{20} - 28.3^\circ$  (c, 1.8, water); lit.<sup>11</sup>: m.p. 179-180°,  $[\alpha]_D^{20} - 27^\circ$ ; <sup>13</sup>C-n.m.r. spectrum identical to that reported by Bhattacharjee et al.<sup>12</sup>.

In a separate experiment, the amount of 1 present in the hot-water extract, that remained glycosically bound was determined: Hydrolysis with 25mM H<sub>2</sub>SO<sub>4</sub> for 1 h at 80° released only 0.46  $\mu$ mol of additional 1 per ml. In addition, the high-molecular-weight material obtained from the XM-50 concentrate, which should have been similar to the Collocalia mucoid prepared by the method of Howe et al. <sup>13</sup>, was treated with neuraminidase in order to determine whether the material still had any bound sialic acid. No free 1 was detected under a variety of assay conditions. Therefore, it can be concluded that, in the hot-water extract, more than 95% of 1 is present as the free sugar. The chemical nature of Peak II from the ion-exchange chromatography remains to be established; however, no Warren positive material was detected before or after acid hydrolysis (see conditions just described), and the  $R_F$  (on t.l.c.) of this material was less than that of 1.

This simplified isolation procedure at a minimal cost and with few manipulations provides substantial quantities of 1 which is useful for further synthetic purposes.

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